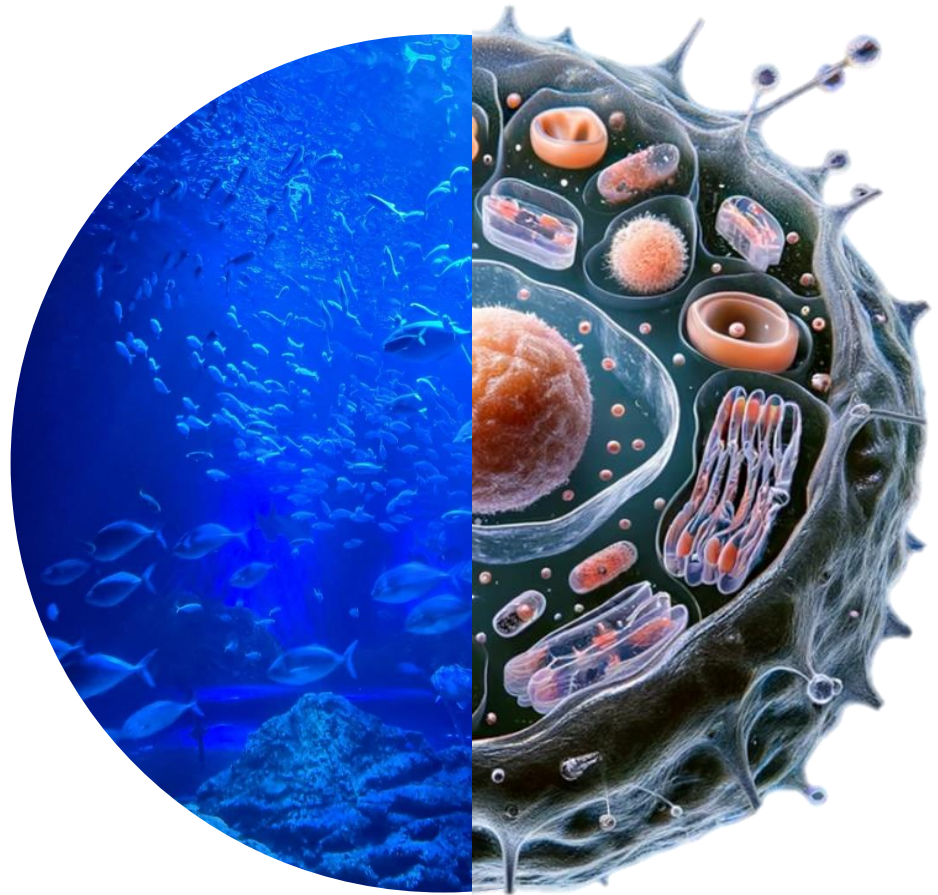




# Single cell sequencing is the future of population genetics in marine environments.

Haylea Power  
Environomics Future Science Platform  
CSIRO  
21<sup>st</sup> February 2025





# Genomic analyses from environmental samples

We all do Environmental DNA (eDNA) studies

- ✓ We already do community diversity surveys or species-specific assays with eDNA samples  
... but can we do more with eDNA samples?

- ❑ The challenge is obtaining more comprehensive genetic information from eDNA samples which previously required tissue samples.
  - ❑ Multi-locus genotypes to identify individuals
  - ❑ Measuring animal abundance
  - ❑ Population dynamics

***Why haven't we done this yet?***





# eDNA limitations



**Unsorted genetic material:**

Genomes and fragments from multiple species and individuals.



**Highly heterogeneous and degraded:**

Material includes degraded tissue to <100 bp fragments.



**Limited focus on the physical characteristics of collected material:**

Perhaps because current methods seem effective?



***Can we improve on this?***





# eDNA +

## Beyond presence-absence: Improving eDNA performance

Exploring the physical components of eDNA samples. Can we:

- ❑ Physically separate components of eDNA to target taxa?
- ❑ Target components of eDNA with fluorescence activated cell sorting (FACS)?
- ❑ Enrich eDNA samples without inhibitors and non-target taxa (i.e. bacteria)?
- ❑ Enable shotgun or long read sequencing of “cleaner” eDNA samples to retrieve richer genomic information?

### The aim:

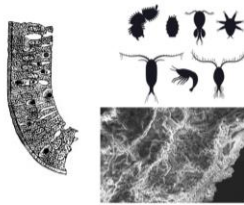
Develop a non-invasive method to obtain richer information from environmental samples



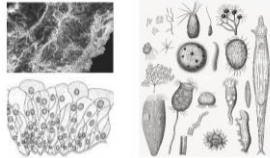


### Physical components of eDNA sample

Whole organisms, tissue and biofilm aggregates



Smaller tissue and biofilm aggregates, cells, large organelles



Eukaryotic and microbial cells, organelles, chromosomes



Microbial cells, organelles, cellular debris, chromosomes and fragments



Cellular debris, chromosomes and fragments



Cellular debris, fragmented chromosomes and extracellular DNA



Extracellular DNA



80  $\mu\text{m}$

10  $\mu\text{m}$

5  $\mu\text{m}$

1.2  $\mu\text{m}$

0.45  $\mu\text{m}$

0.22  $\mu\text{m}$

FFT

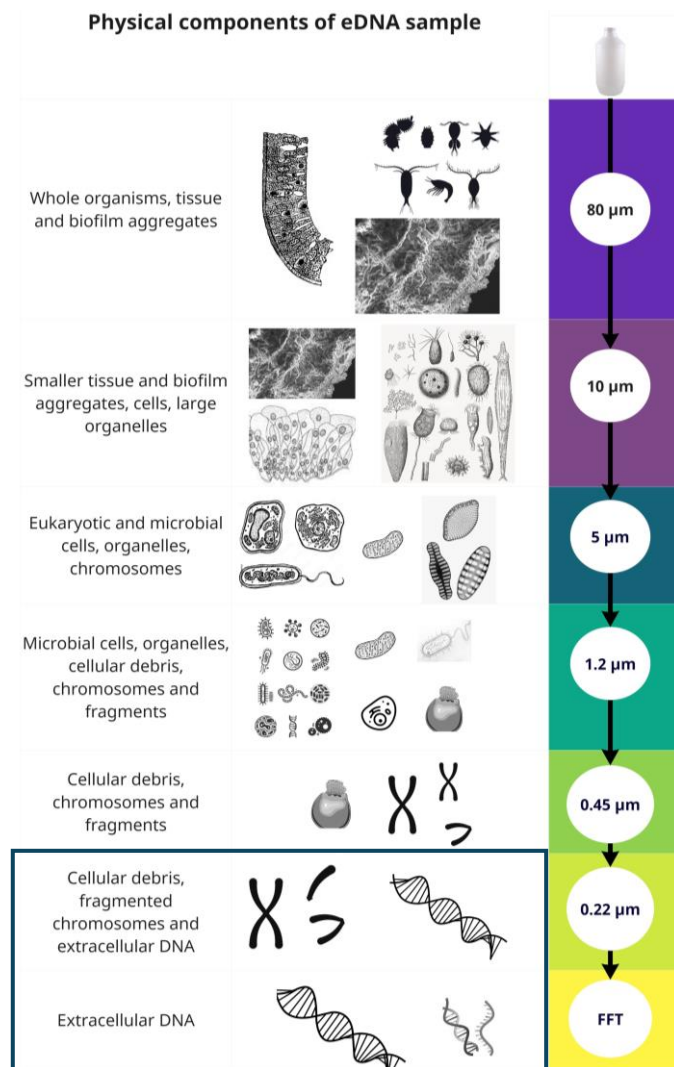
# So, What is eDNA?

## eDNA components hypothesis





# eDNA components hypothesis



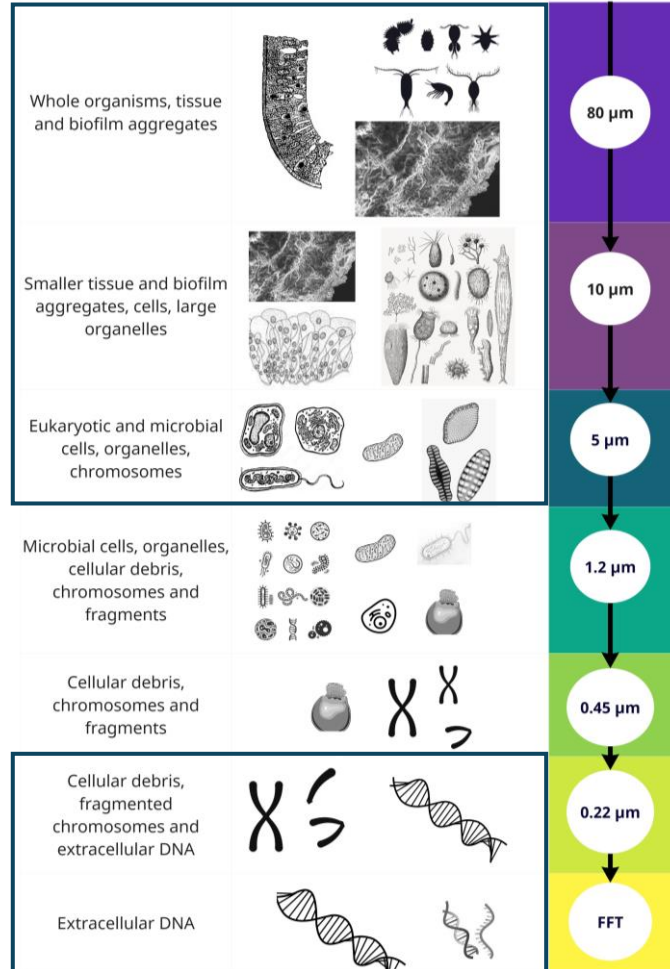
← What people think they're collecting





### Physical components of eDNA sample

## eDNA components hypothesis



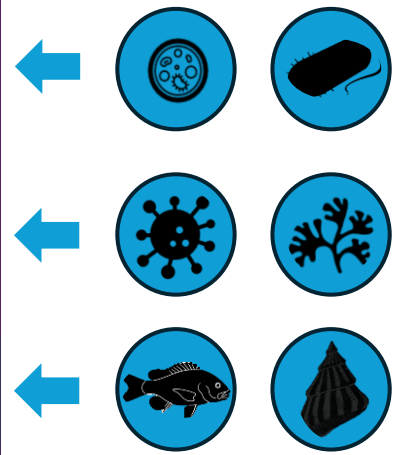
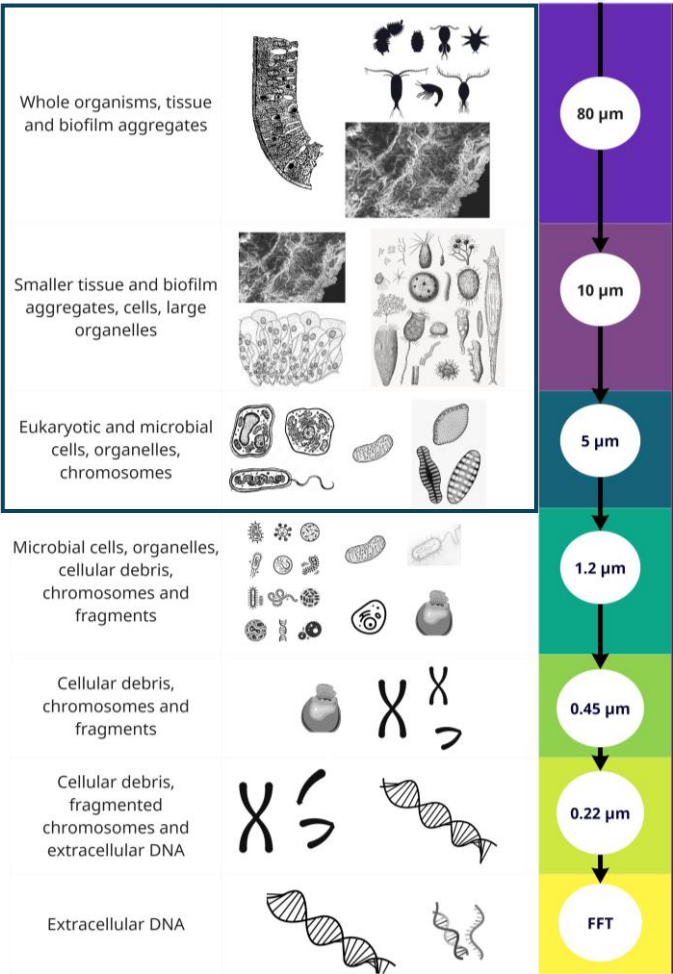
← Where the high-quality DNA is for sequencing depending on target

← What people think they're collecting





### Physical components of eDNA sample



## eDNA components hypothesis

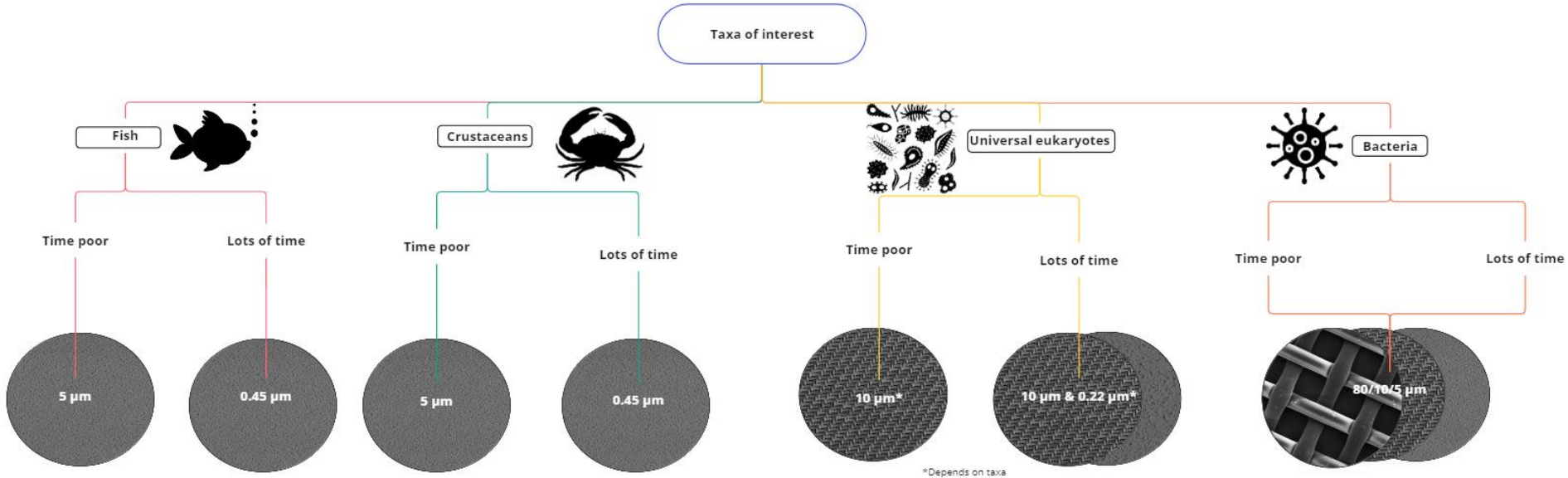
Where the high-quality DNA is for sequencing depending on target





# How to enrich your eDNA samples

## How to sample eDNA for your desired taxa





# Why does this matter?

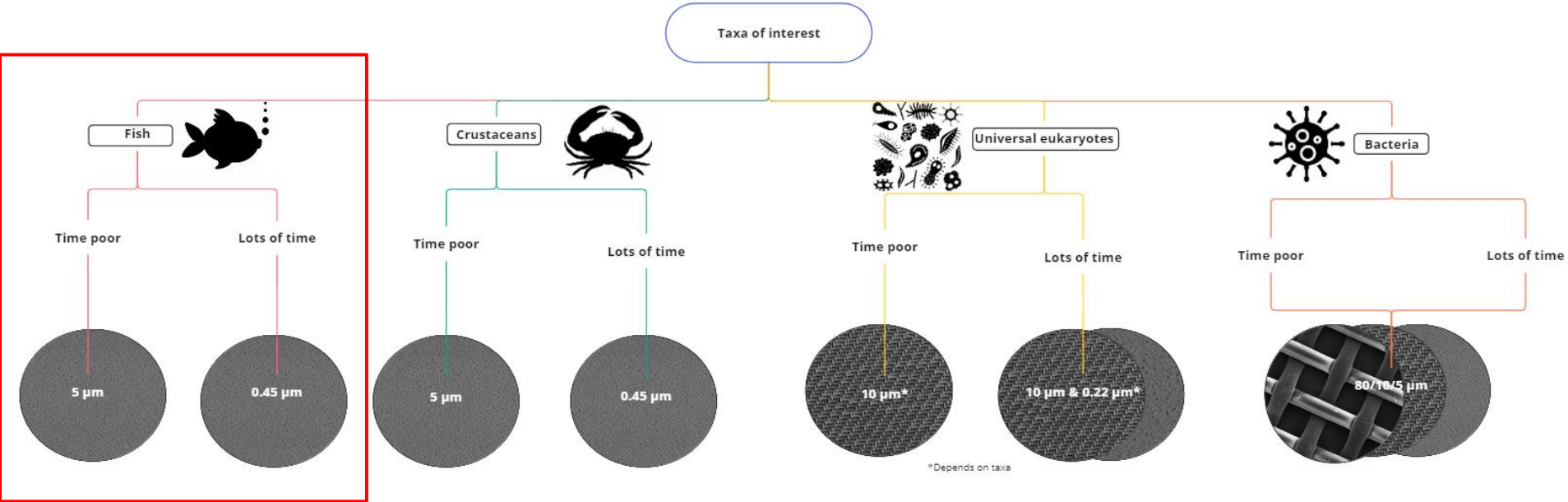
- Can we physically separate components of eDNA to target taxa? (aka enriched eDNA sample)
  - ✓ YES!
- Now, can we target components of eDNA with fluorescence activated cell sorting (FACS)?





# Why does this matter?

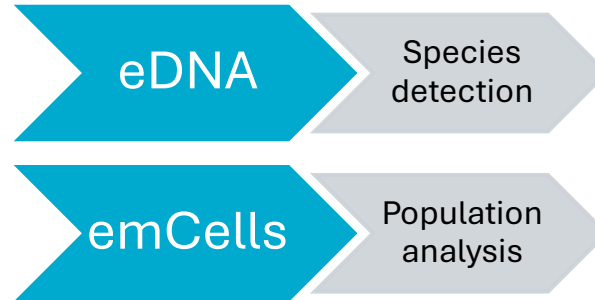
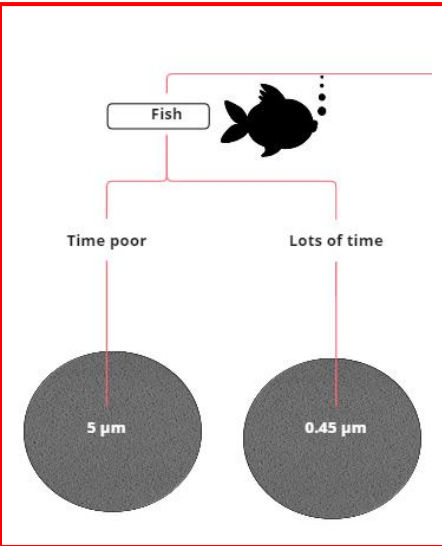
## How to sample eDNA for your desired taxa





# Beyond presence-absence

- Can we use just the cellular components?  
***emCells*** - *Environmental metazoan cells*





# How do we get them?



FACS can sort millions of cells per minute

1. Water collection and cell processing

2. Fluorescence-activated cell sorting (FACS)

3. DNA amplify and sequence single cells

4. Obtain more informative markers for population genetics

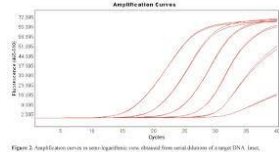
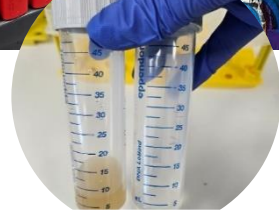
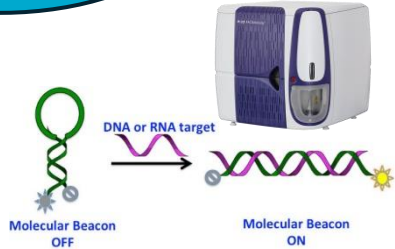
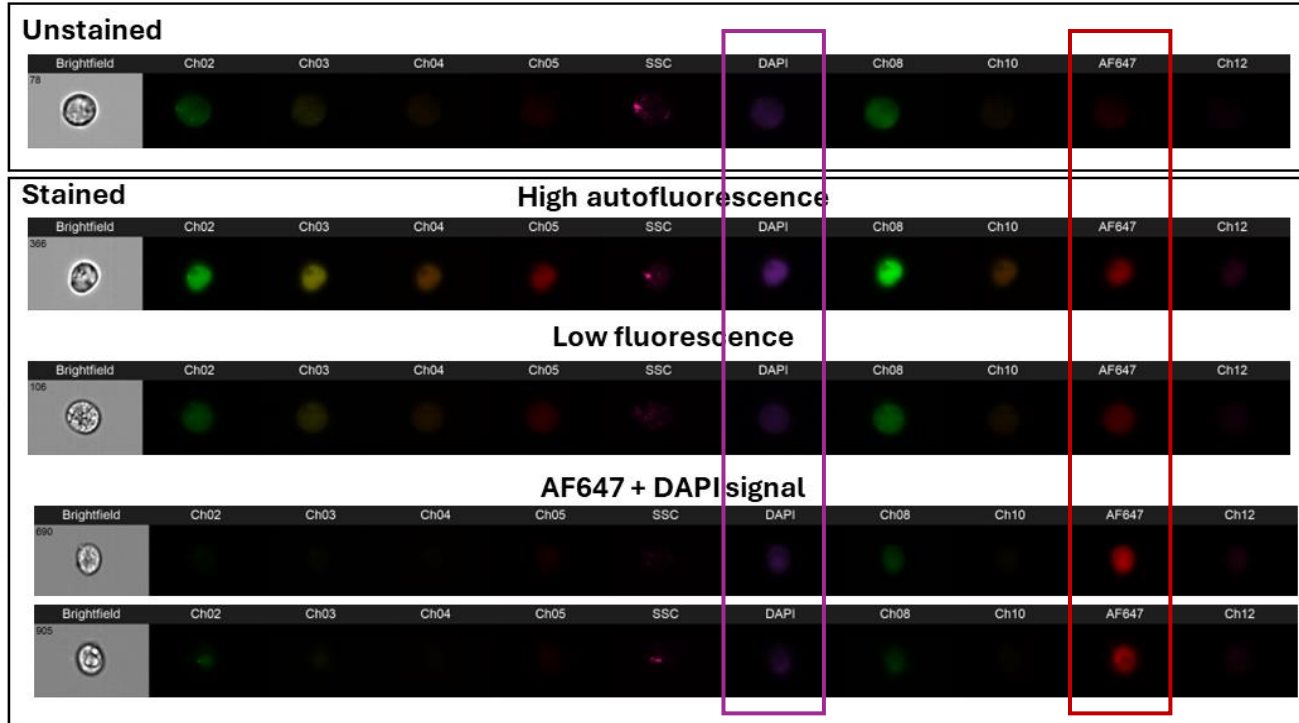


Figure 2. Amplification curves to assess quantitative PCR showed from serial dilutions of target DNA. Lines represent cycle of Amplified DNA, C<sub>t</sub> value.





# Imaging flow cytometry



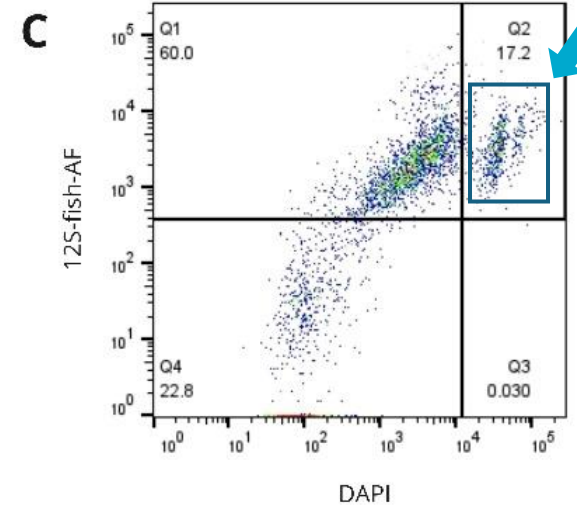
DAPI and probe positive without strong autofluorescence signals





# emCell FACS

Concentrated fish cells

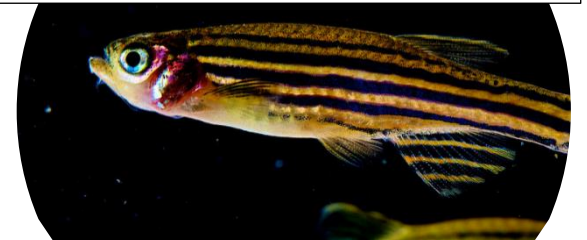
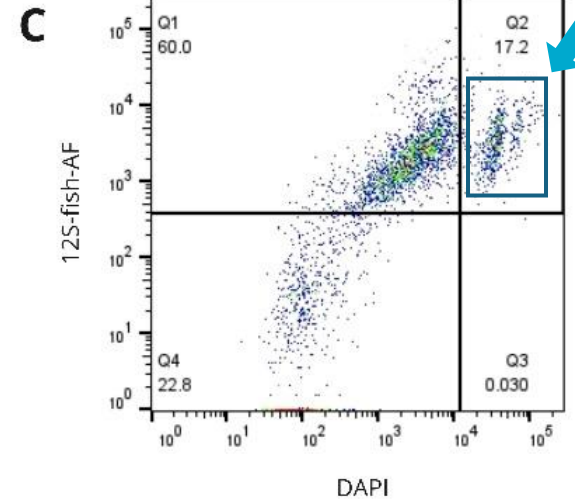
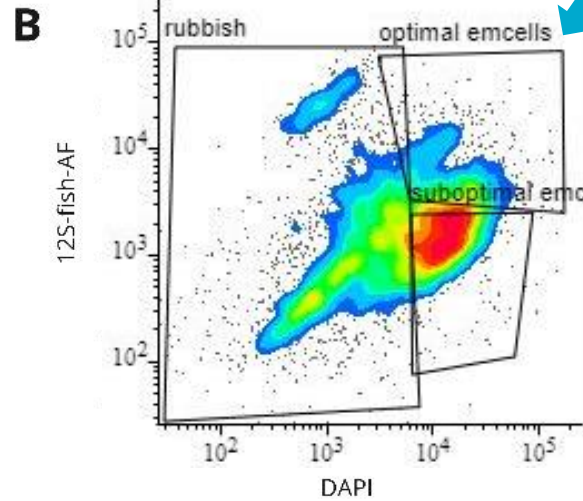




# emCell FACS

Aquarium water

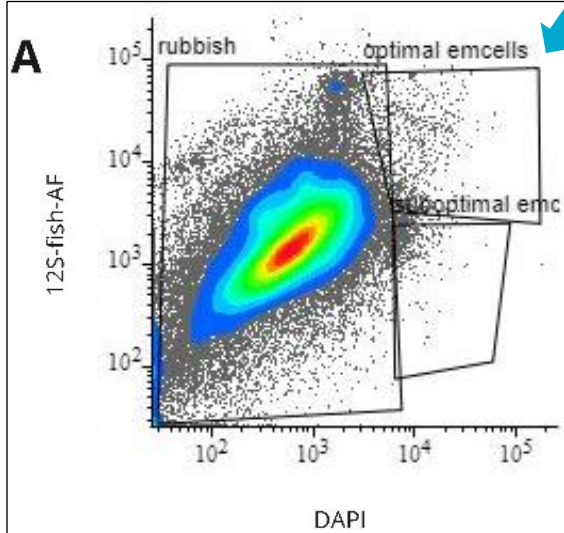
Concentrated fish cells



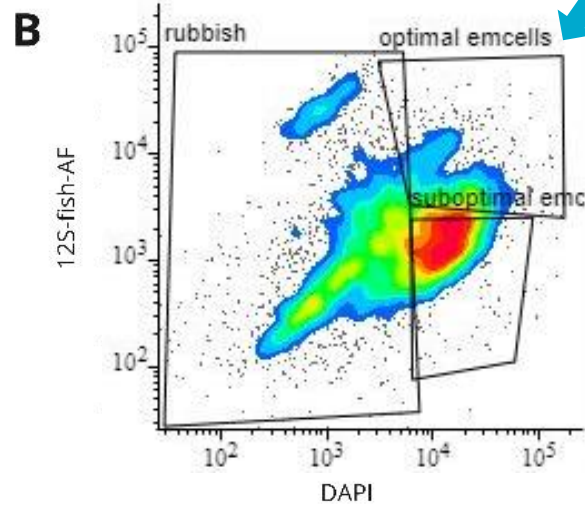


# emCell FACS

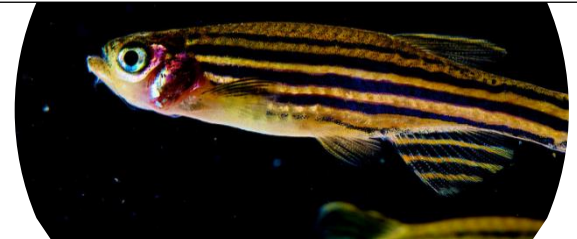
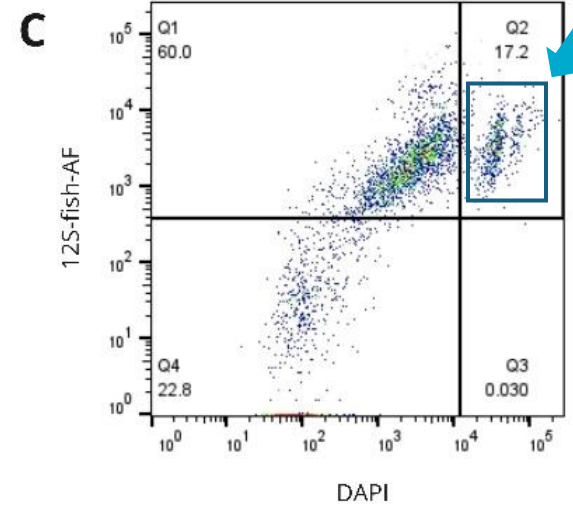
Marine water



Aquarium water



Concentrated fish cells



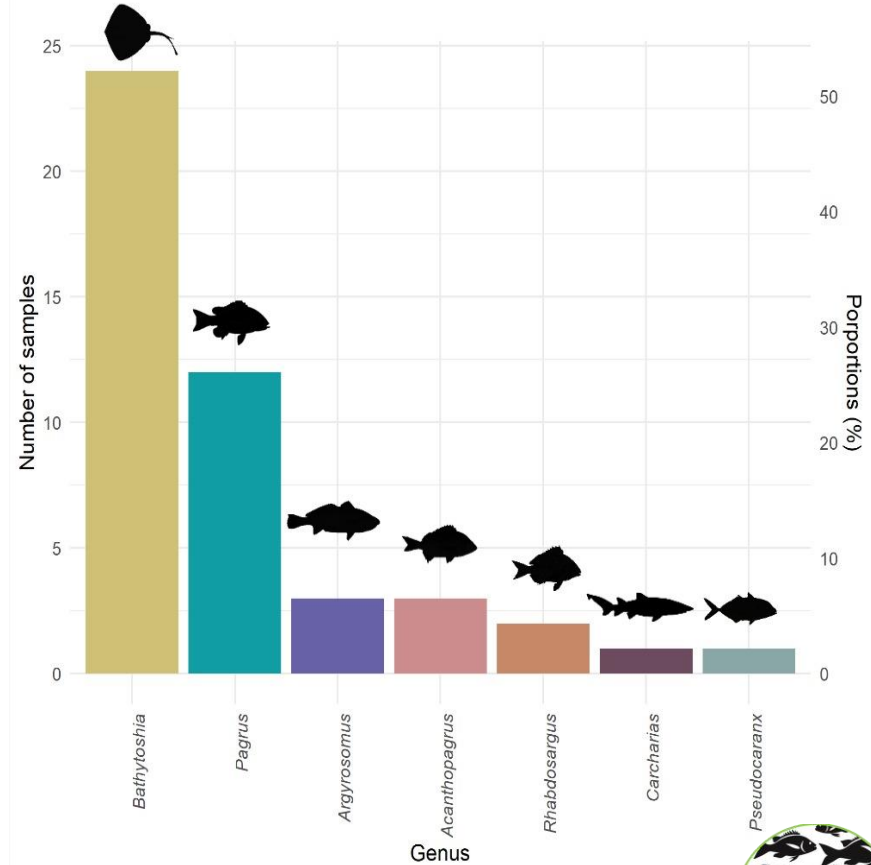


# Isolating emCells



## Aquarium of Western Australia (AQWA)

- 3 million L mesocosm
- 38 species of bony fish, sharks and rays
- Two targets: 16S and 18S rRNA
- 25% (n = 49) success rate



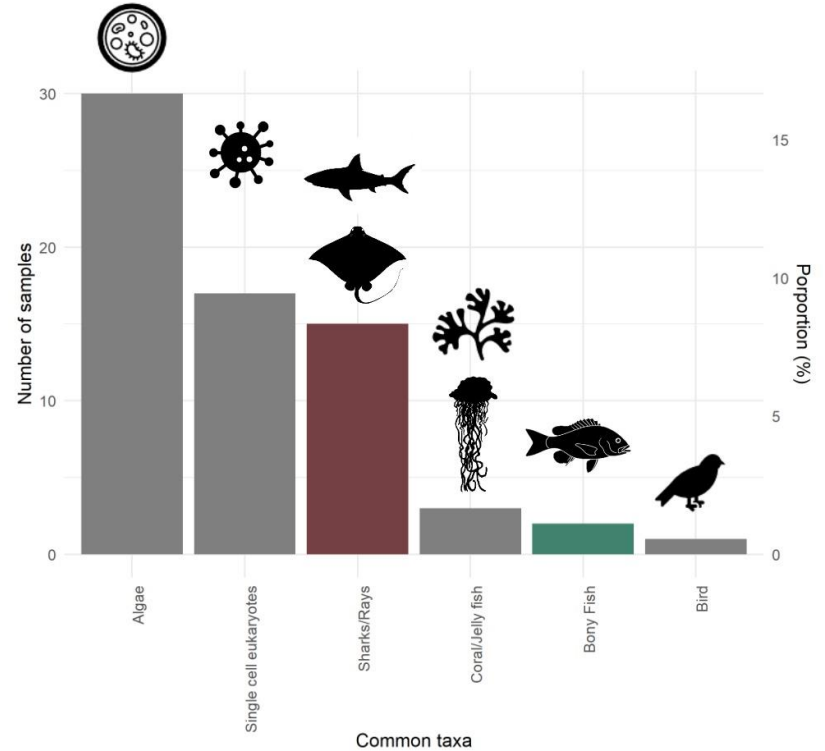


# Off-targets

Autofluorescent cells in the same size range

- **Algae!**
- Single cell eukaryotes
- Coral and jellyfish
- Birds

Working on increasing specificity



B)





# Single cell analyses

Once you have enriched eDNA samples or emCells you can do:

- Multilocus genotyping
- Individual identification
- Mark-recapture population abundance
- Gene flow
- Population structure
- Migrant detection
- Sex ratios
- And more.....





# Single cell analyses

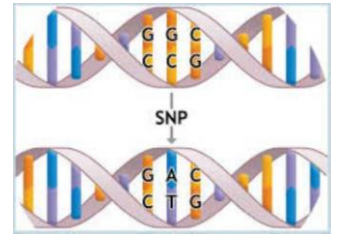
Once you have enriched eDNA samples or emCells you can do:

- Multilocus genotyping
- Individual identification
- Mark-recapture population abundance
- Gene flow
- Population structure
- Migrant detection
- Sex ratios
- And more.....





# Population genetics 101



- SNP's (Single Nucleotide Polymorphisms)
- Can use to identify individuals
- Normally need tissue samples
- Difficult from eDNA since they are not linked
- Single cells maintain these linkages





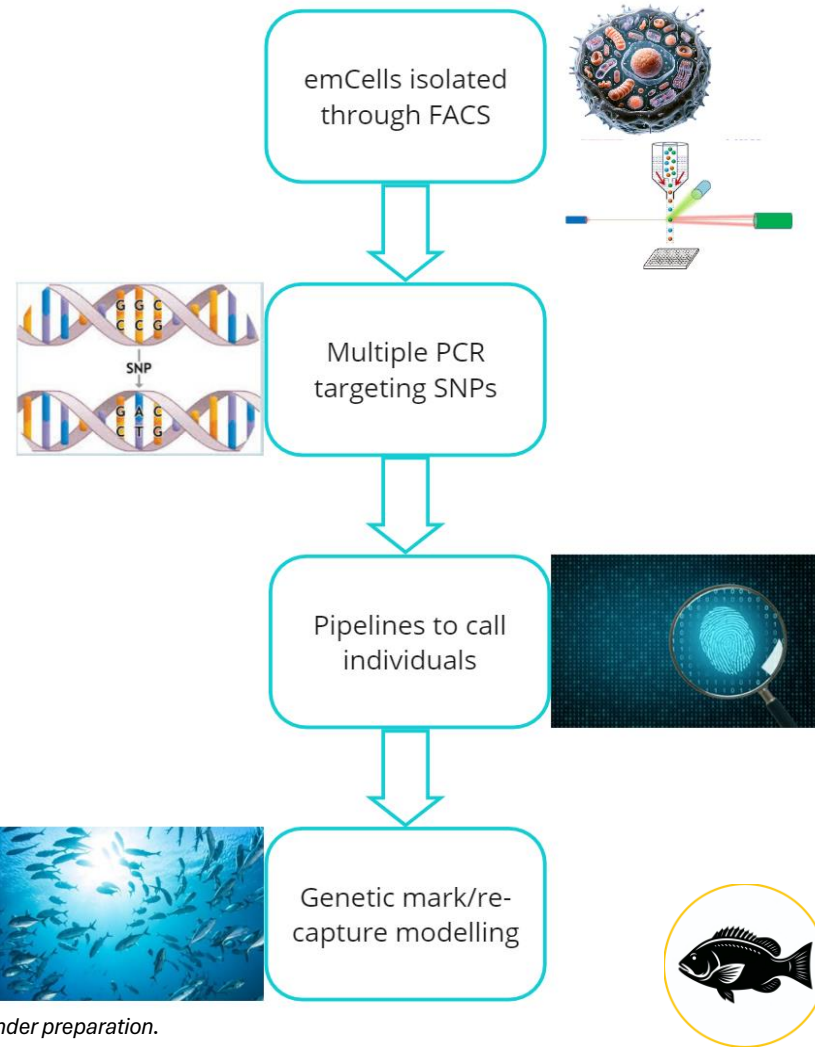
# Single cell SNP analysis

## Experimental set up

- Tissue samples and single cells
- SNP assay
- Bioinformatic pipeline

## Results

- >91% successful
- Minimum 60% allele coverage needed

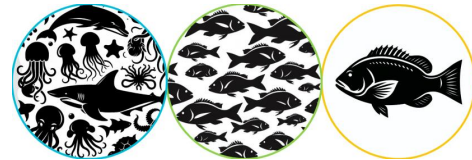




# eDNA + Beyond presence-absence

- We can get more from eDNA that just presence-absence!
- Since we can enrich eDNA samples now, we can
  - Reduce background noise
  - Enrich target acquisition
  - Single-cell sequencing is feasible
- Enable non-invasive methods to obtain richer information from environmental samples

***Exciting future potential!***





Thank you



*“Using genomics, bioinformatics and nano-technologies, Environomics is finding new resources in nature and reinventing how we measure and monitor ecosystem health, change and threats” – Olly Berry*

Funding



Translational Cancer Pathology  
Laboratory & Western  
Australian Zebrafish  
Experimental Research Centre



The CMCA - Australian Microscopy,  
Characterisation & Analysis

Collaborations



THE AQUARIUM OF WESTERN AUSTRALIA



Trau Lab



Curtin University

TrEnD Lab