

State of play: the role of bacterial communities in marine eDNA decay

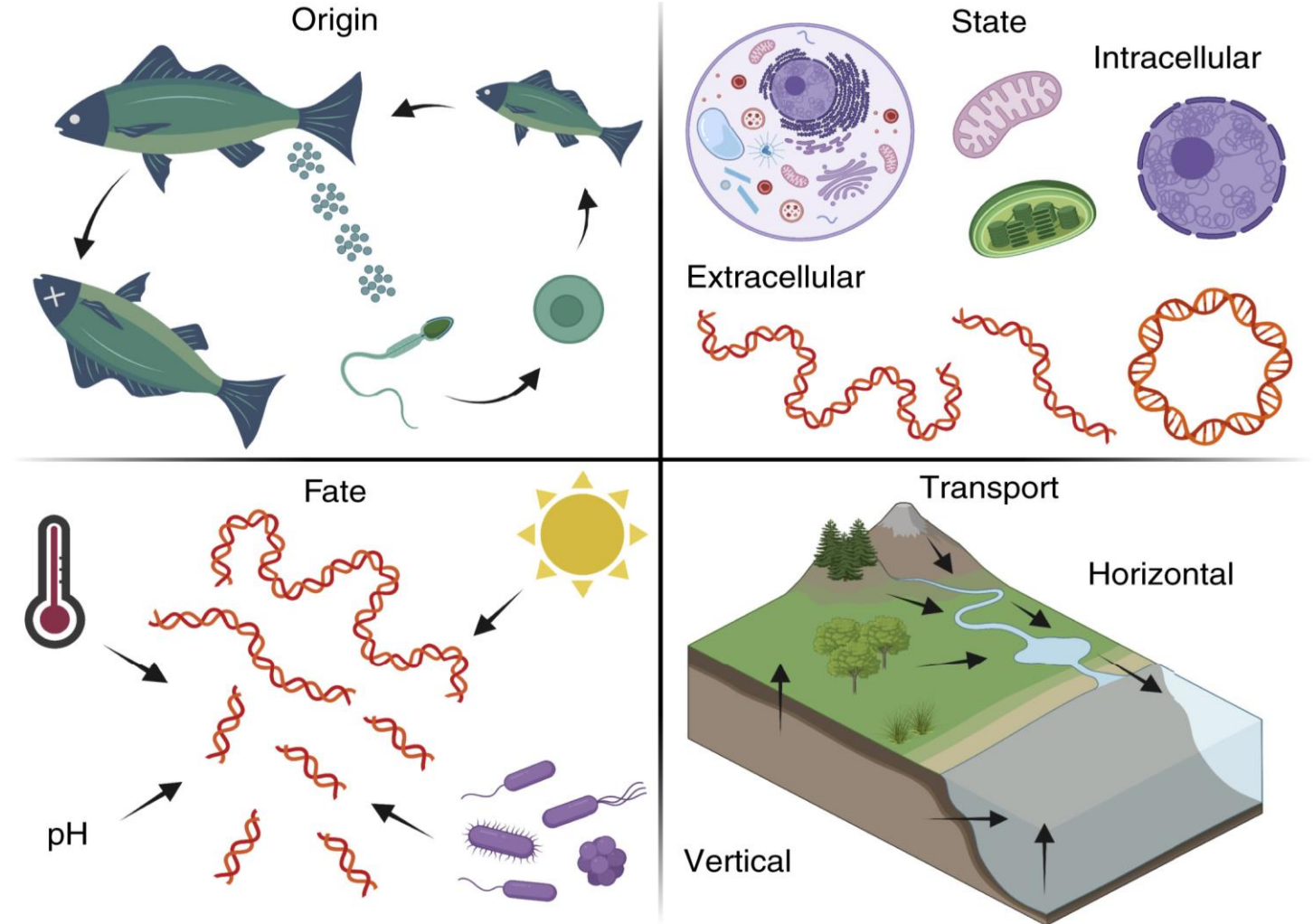
Susanna Theroux, PhD
Southern California Coastal Water Research Project



USC

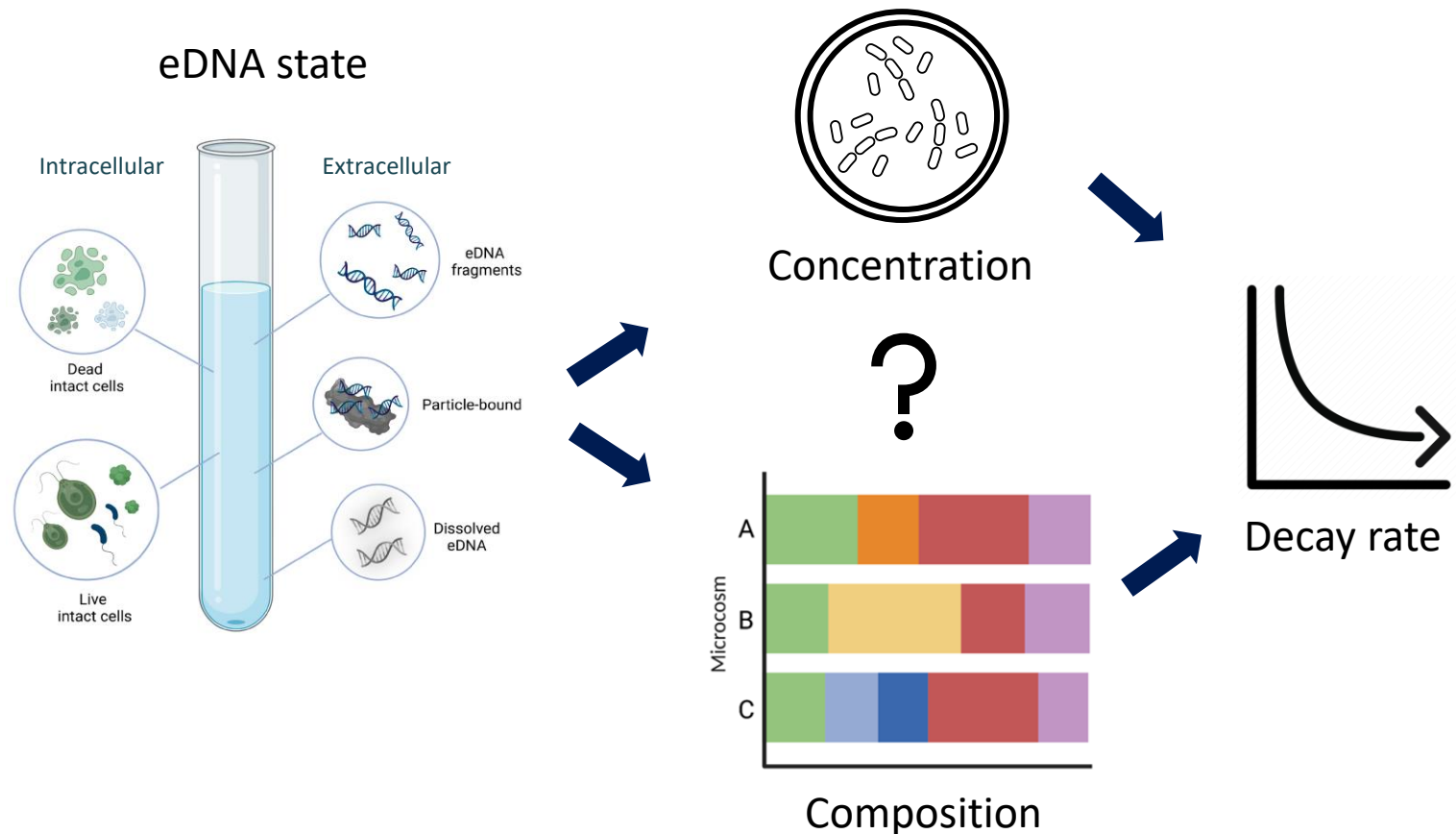
Ecology of eDNA

- Understanding the **ecology of eDNA** helps us better interpret and predict eDNA detections in the wild
- **Shedding, state, fate, and transport** each play a role individually and together
- The **role of bacteria** in decay is known to be important but mechanisms remain poorly understood

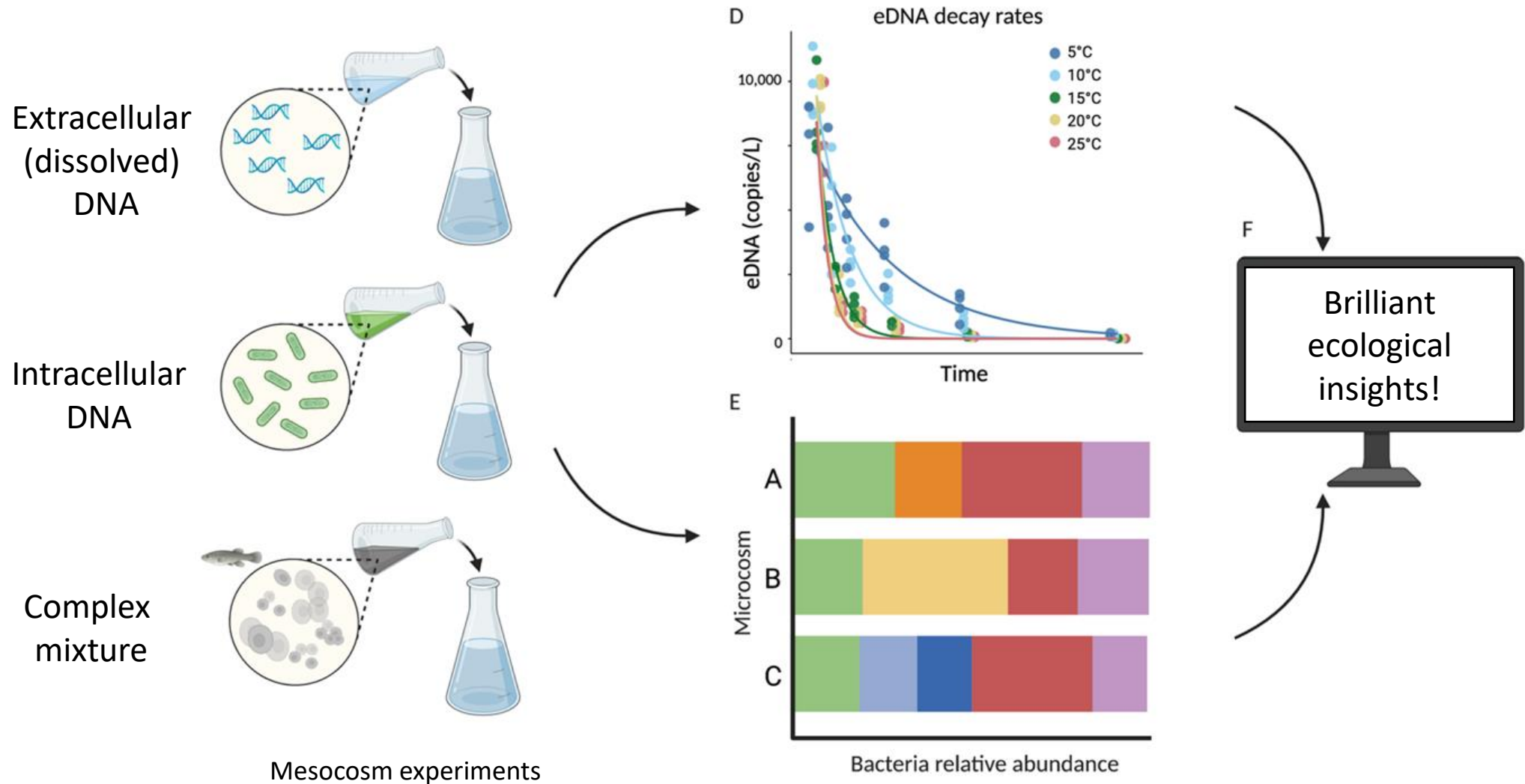


Role of bacteria in eDNA degradation

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Our approach

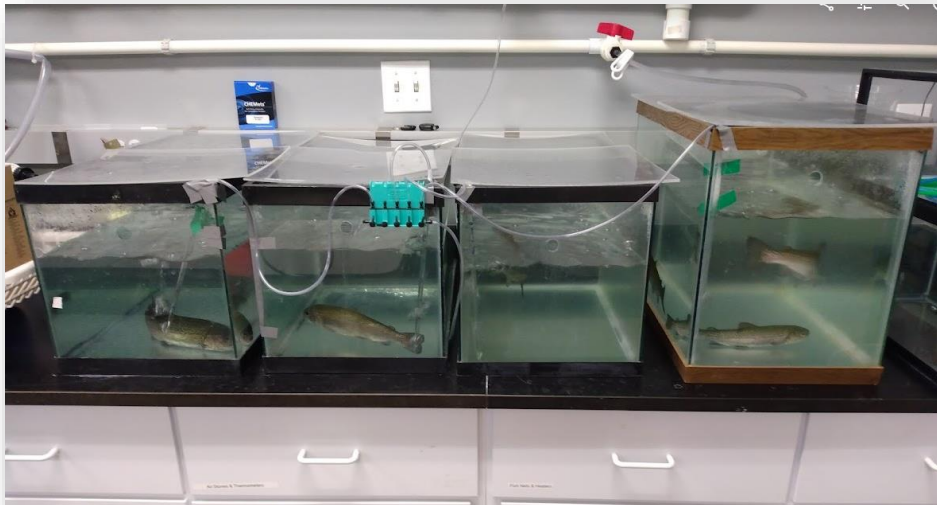


Mesocosm spike-in

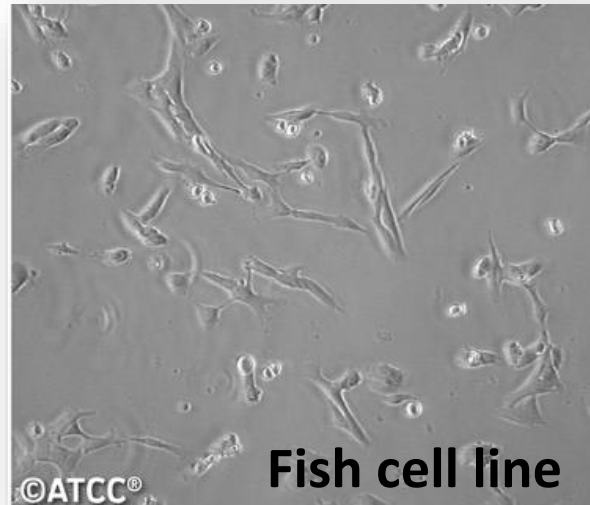
Steelhead / Rainbow trout (*Oncorhynchus mykiss*)



FISH

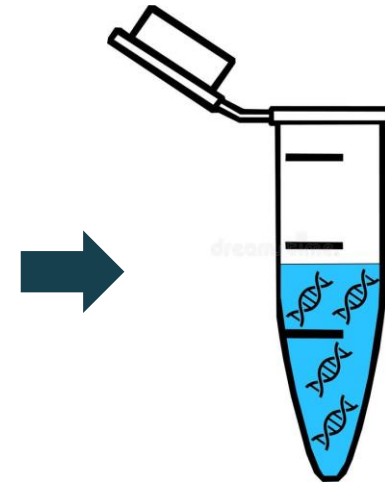


CELL



Fish cell line

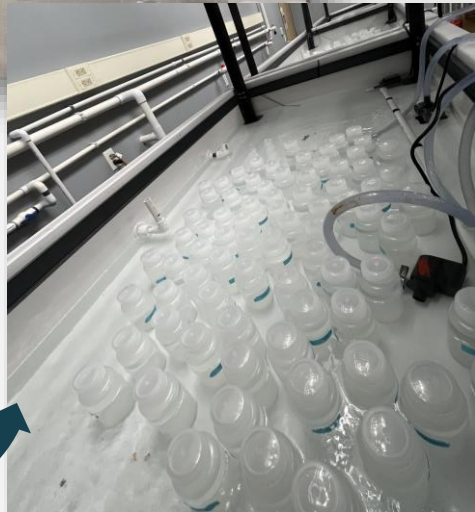
DNA



Mesocosm set-up



Catalina Marine
Protected Area (MPA)
seawater used as a
medium for all
mesocosms



FISH



CELL



DNA

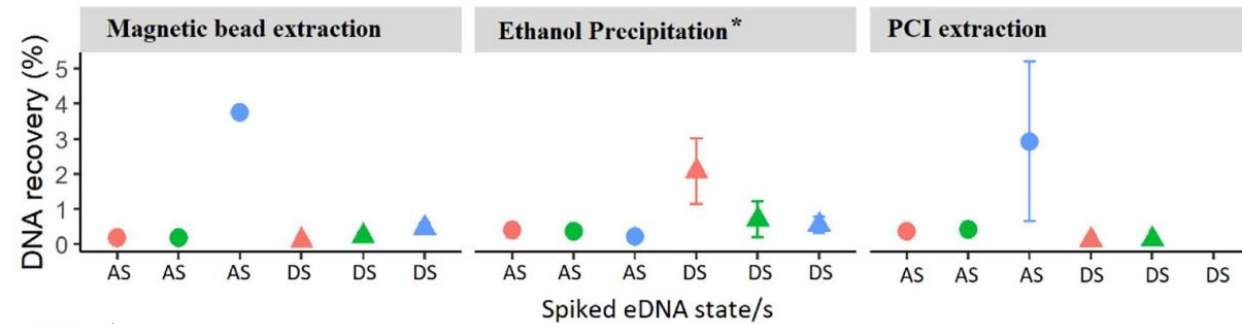


- Three treatments (FISH, CELL, DNA)
- Five temperature (5, 10, 15, 20, 25°C)
- Eight timepoints (0, 4, 8, 12, 18, 24, 48, 72, 168 hours)
- Each treatment in triplicate
- Total = 360 samples

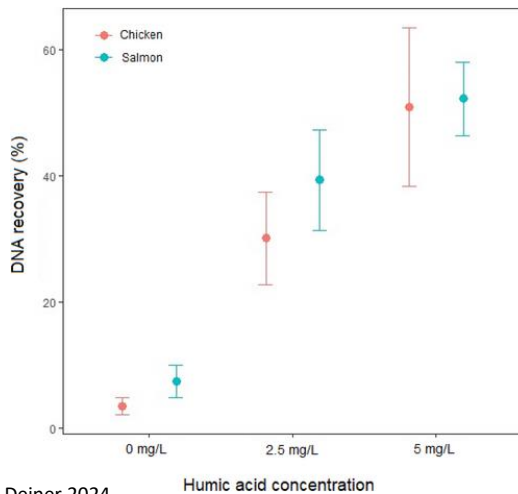
Recovering dissolved DNA

Dissolved DNA is tricky to recover

Chicken DNA target (spiked in free DNA state)



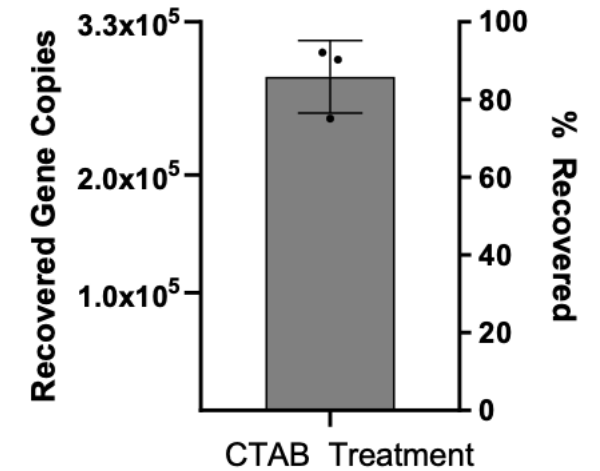
Kirtane et al 2023



Kirtain and Deiner 2024

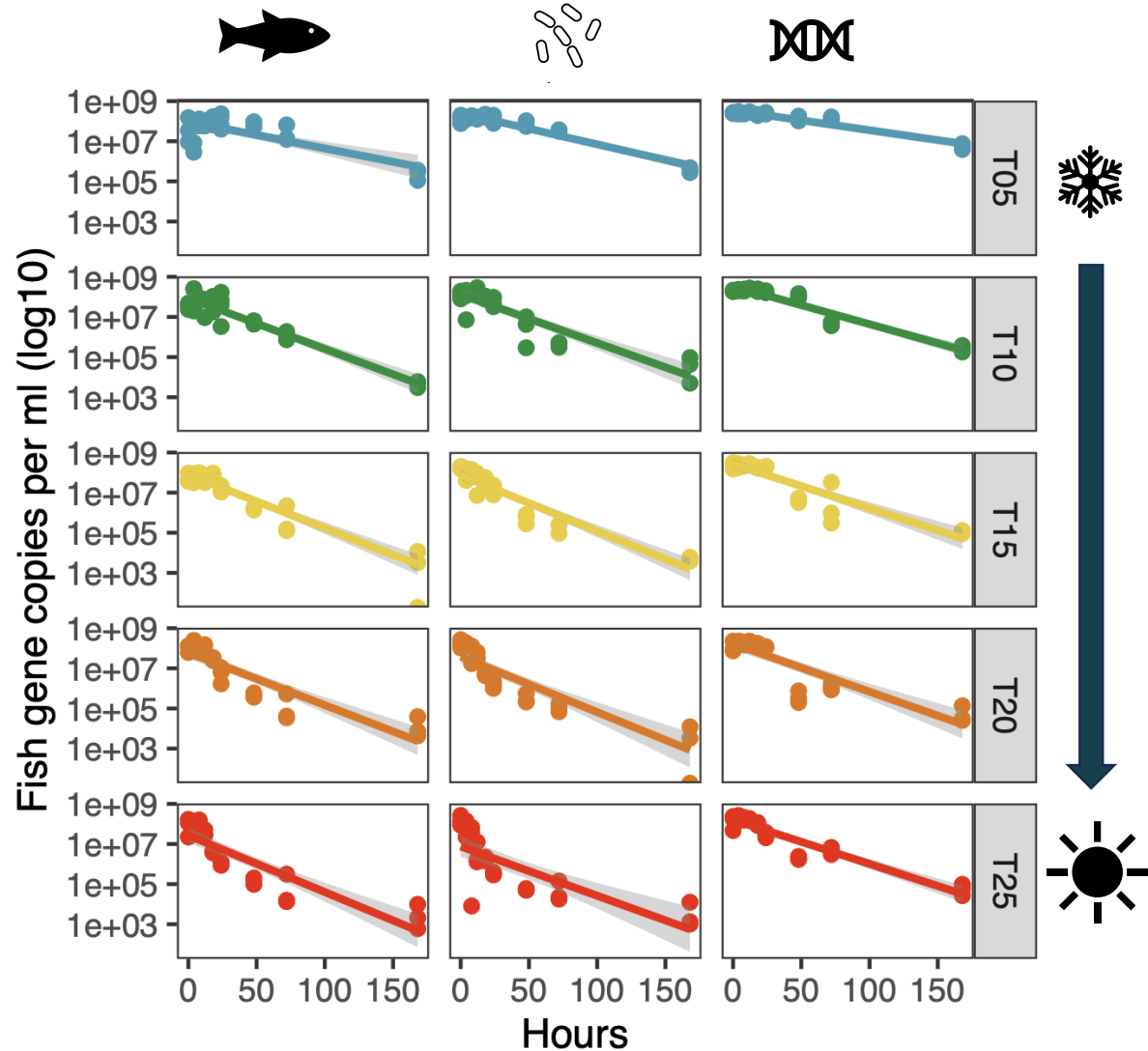


Our modified CTAB method recovered ~90% of dissolved DNA



- Spiked extracted DNA from *O. mykiss* cells
 - Known concentration of target gene copies
- ~80-95% recovery of dissolved DNA with CTAB
 - *O. mykiss* COI ddPCR (Brandl et al., 2015)
- Non-CTAB method recovers ~1% of DNA

eDNA decay rates vary by temperature and state



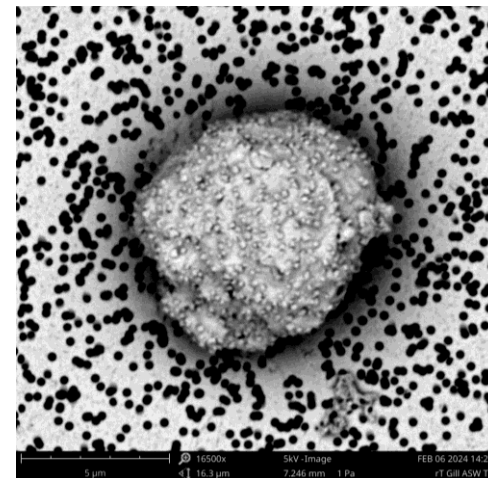
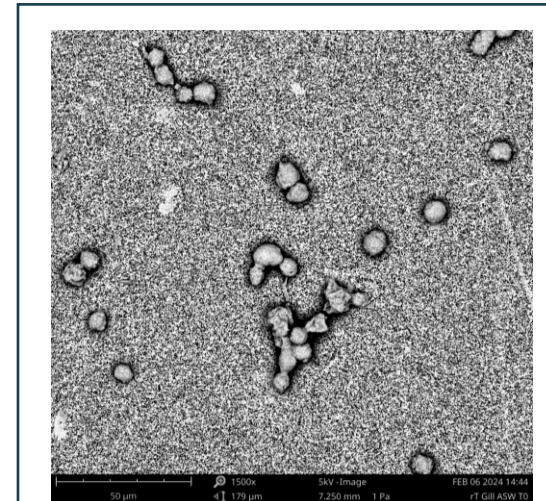
- The expected:
 - Warmer temperature = faster degradation
- The unexpected:
 - “Dissolved” DNA decayed slower than fish and cell DNA
 - At 5°C, DNA $t_{1/2}$ = 67 hours
 - At 5°C, CELL $t_{1/2}$ = 36 hours
 - At 25 °C, DNA $t_{1/2}$ = 10 hours
 - At 25 °C, CELL $t_{1/2}$ = 6 hours

Why did the dissolved DNA degrade slowest?

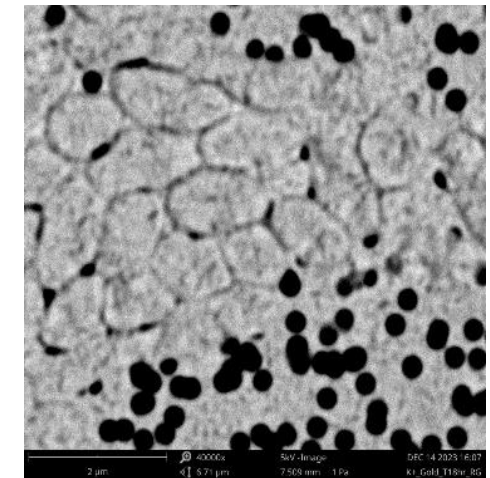
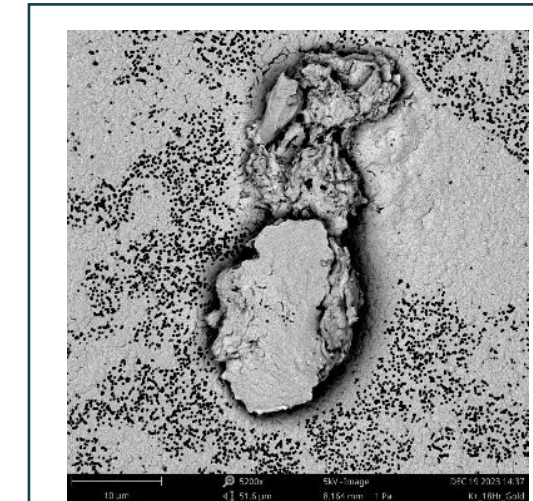
- The FISH and CELL spike-ins were introducing **more organic matter**, which bacteria can readily utilize
- In the dissolved DNA spike-in, the bacteria have **no surface to colonize**
- This may lead to decreased efficiency in DNA degradation and longer decay times

O. mykiss cell line
spiked into seawater

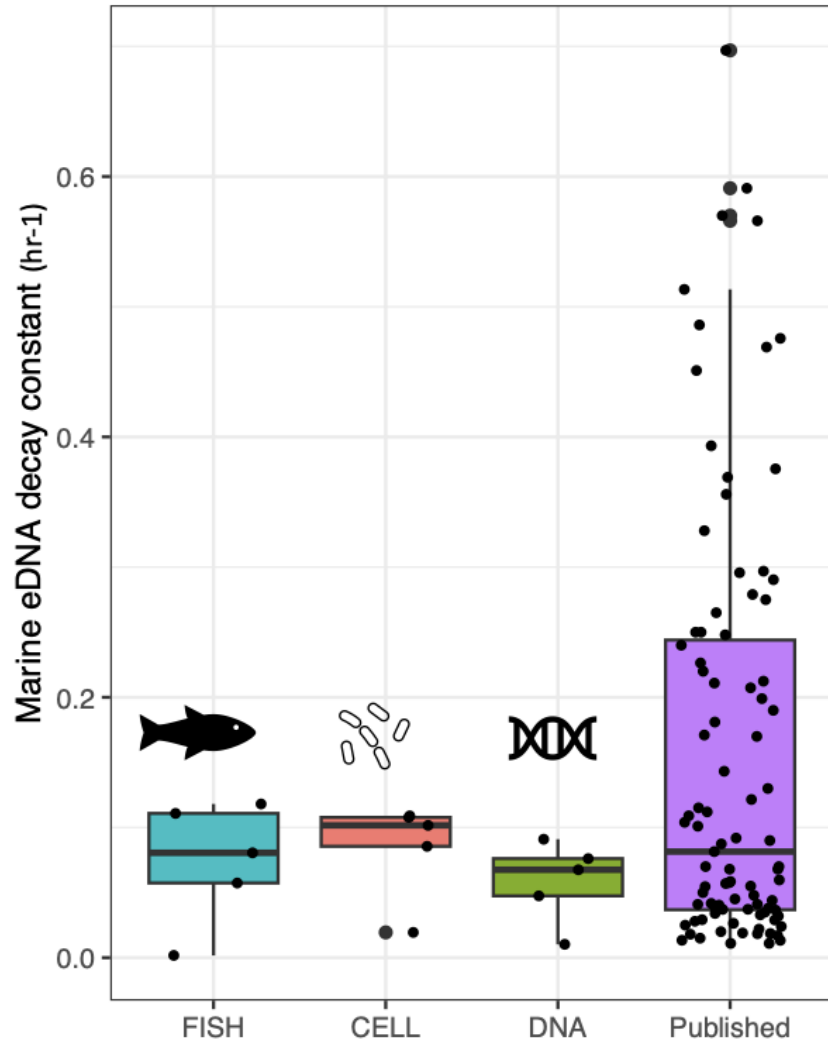
T = 0 hr



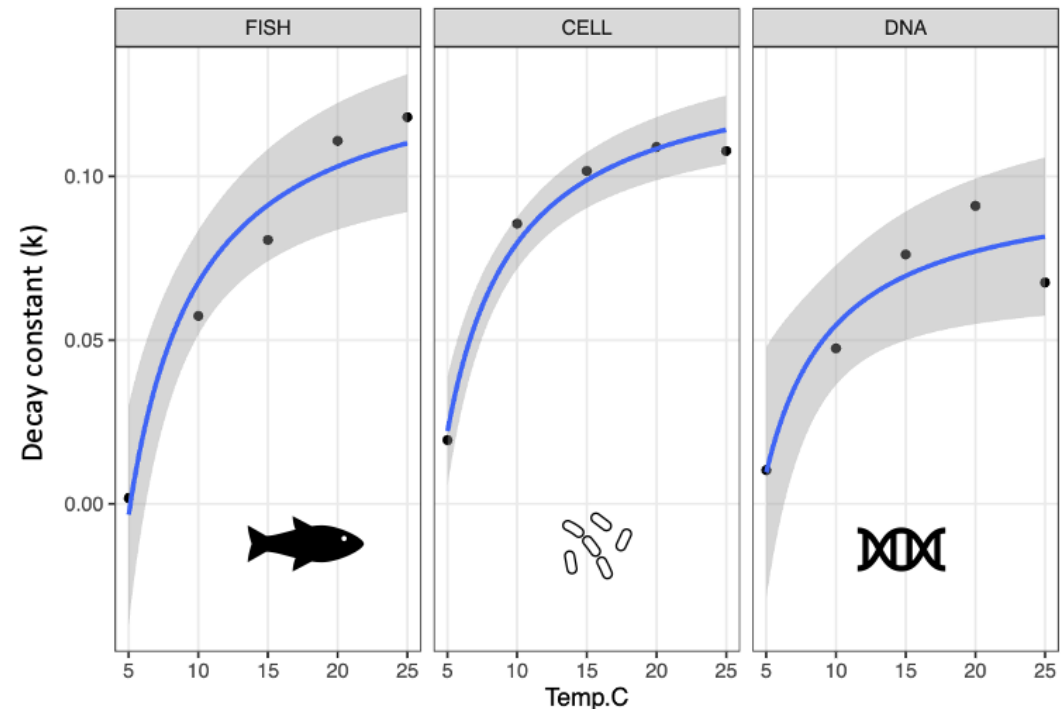
T = 24 hr



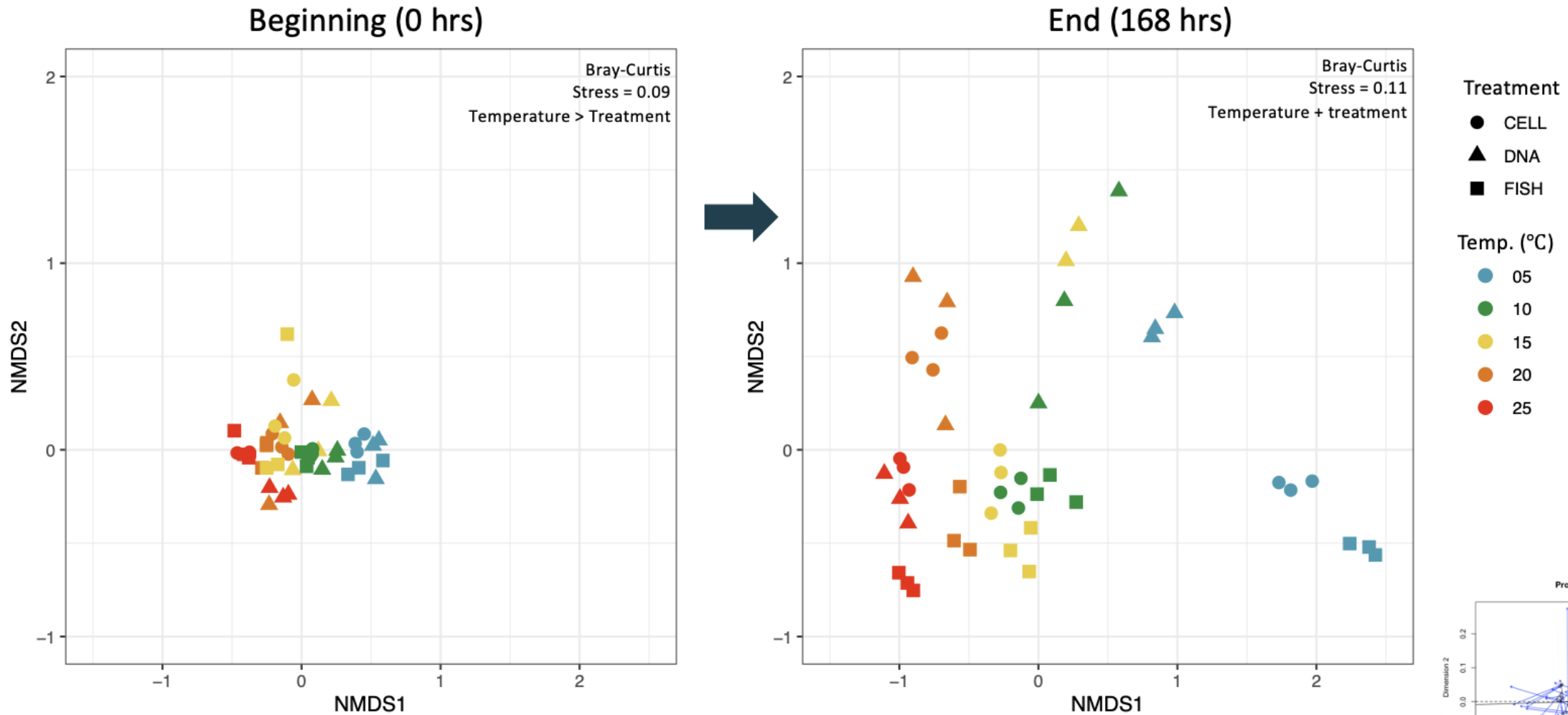
eDNA decay rates vary by temperature and state



- All eDNA states decayed faster at higher temperatures
- “Dissolved” DNA decayed slower than fish and cell DNA

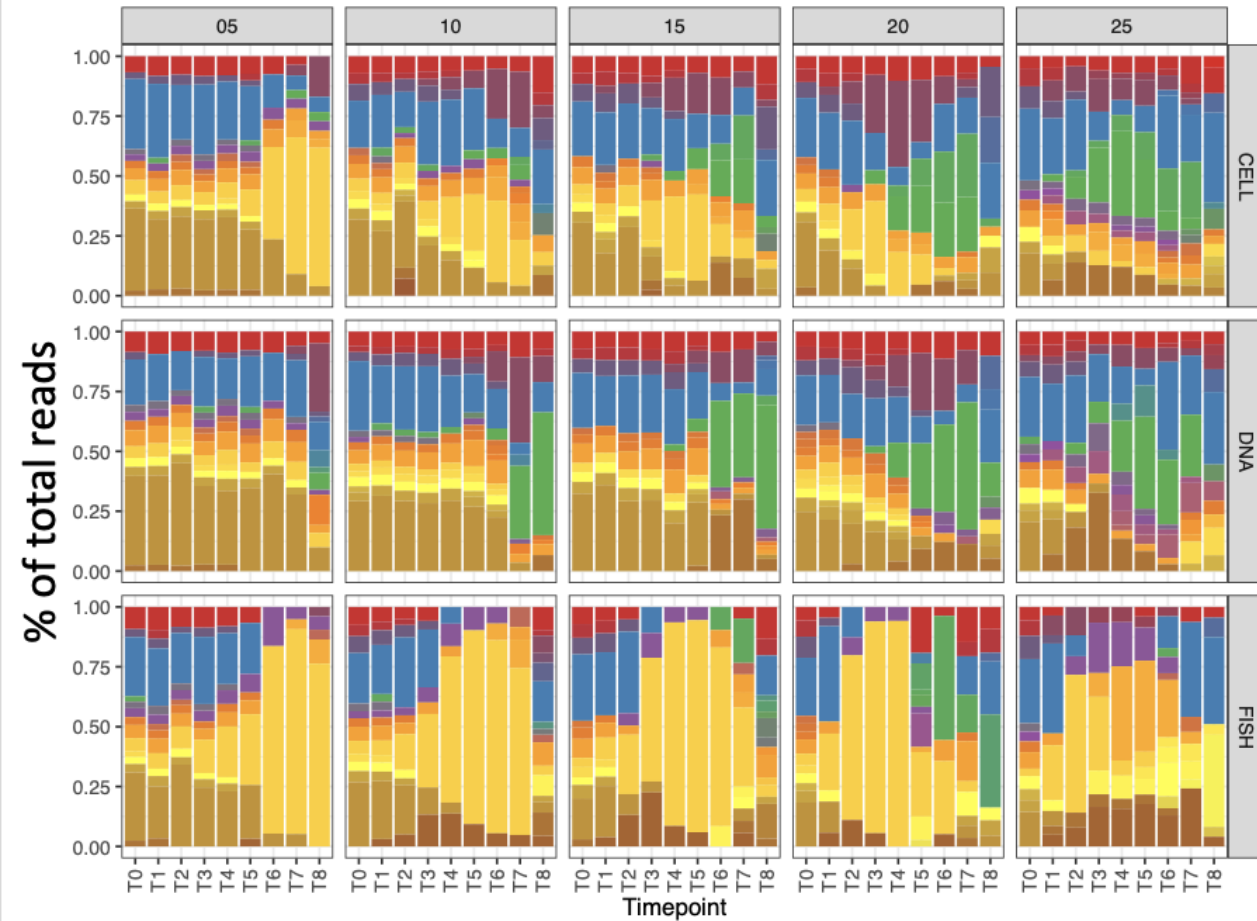


Microbial community changes over time

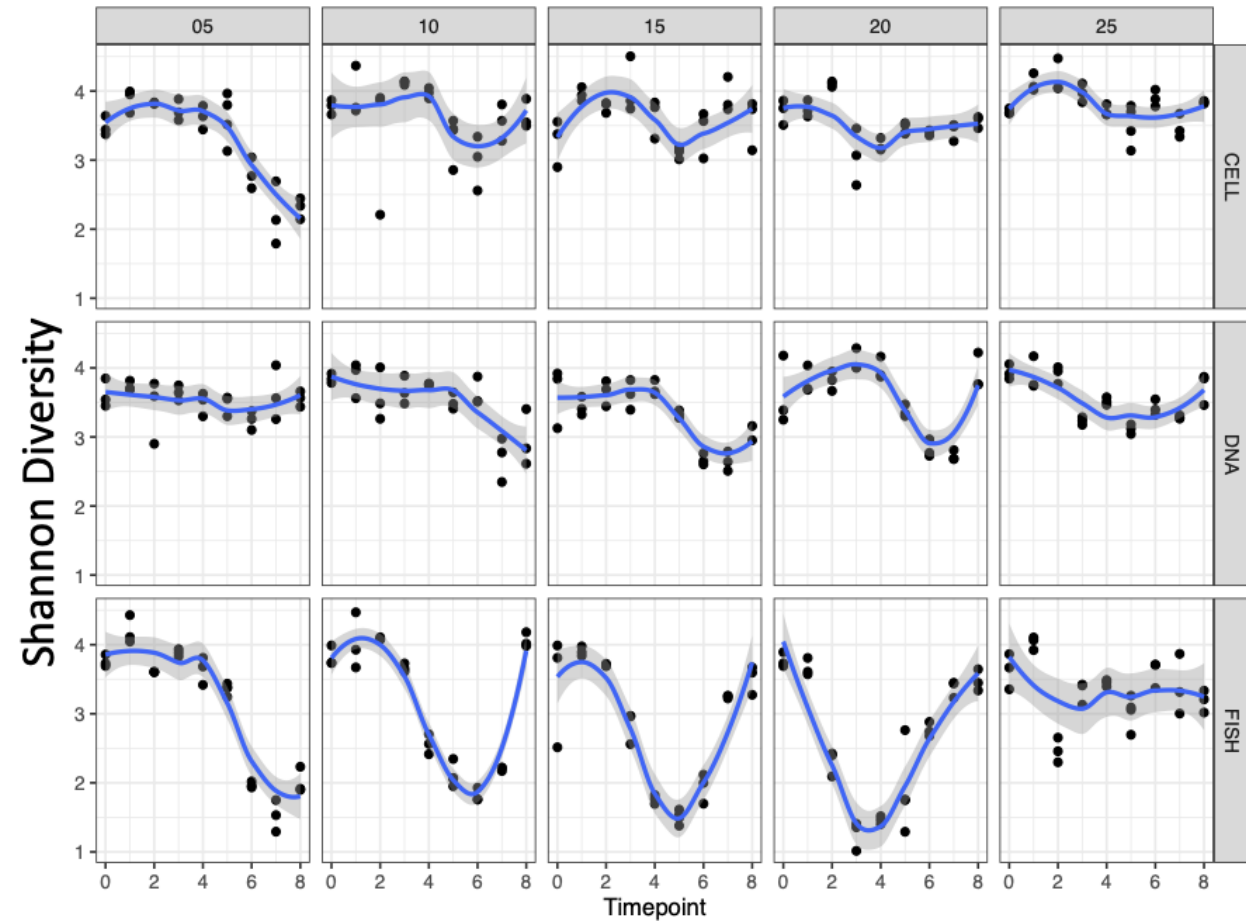


Microbial community changes over time

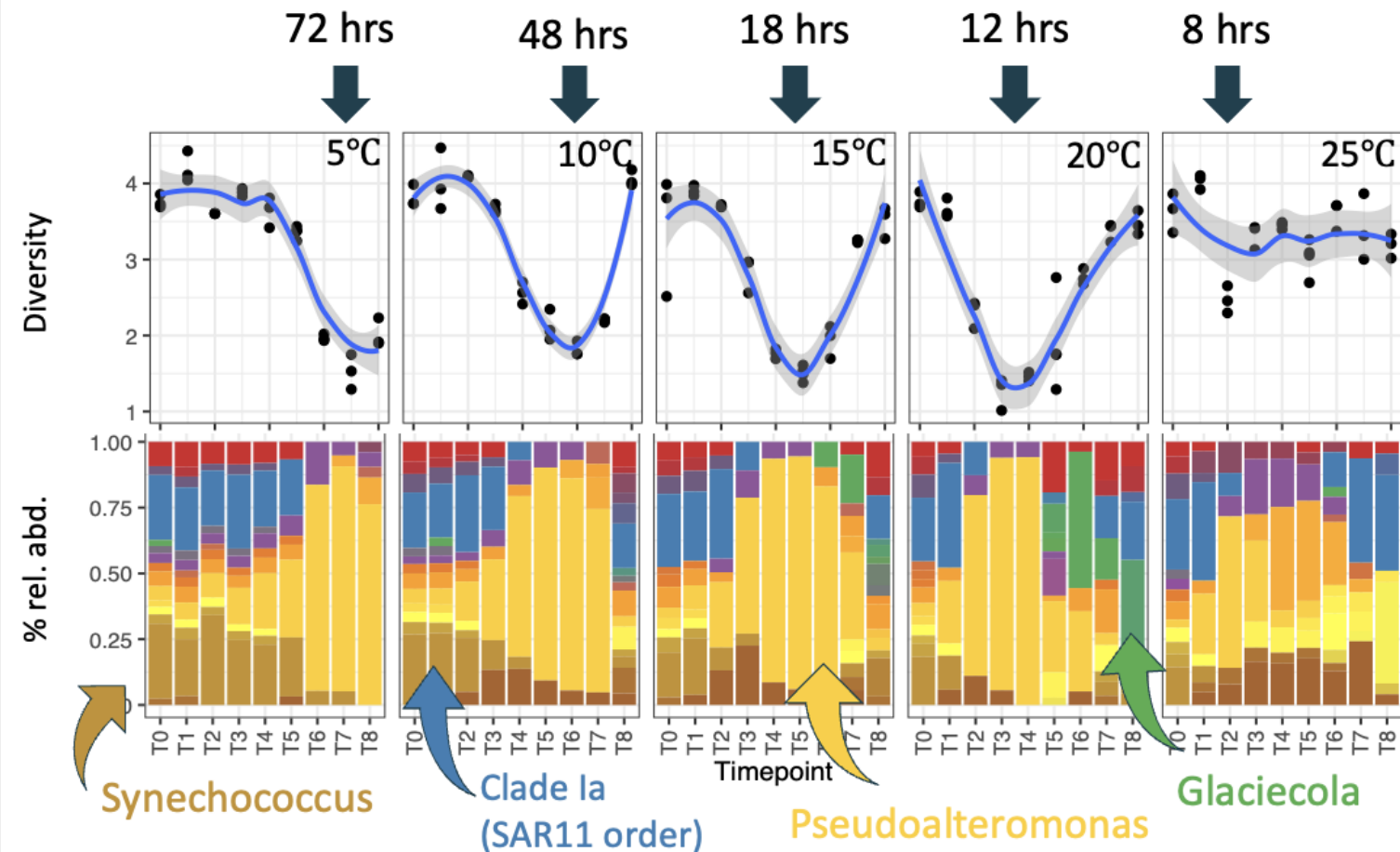
Most abundant genera



Biodiversity

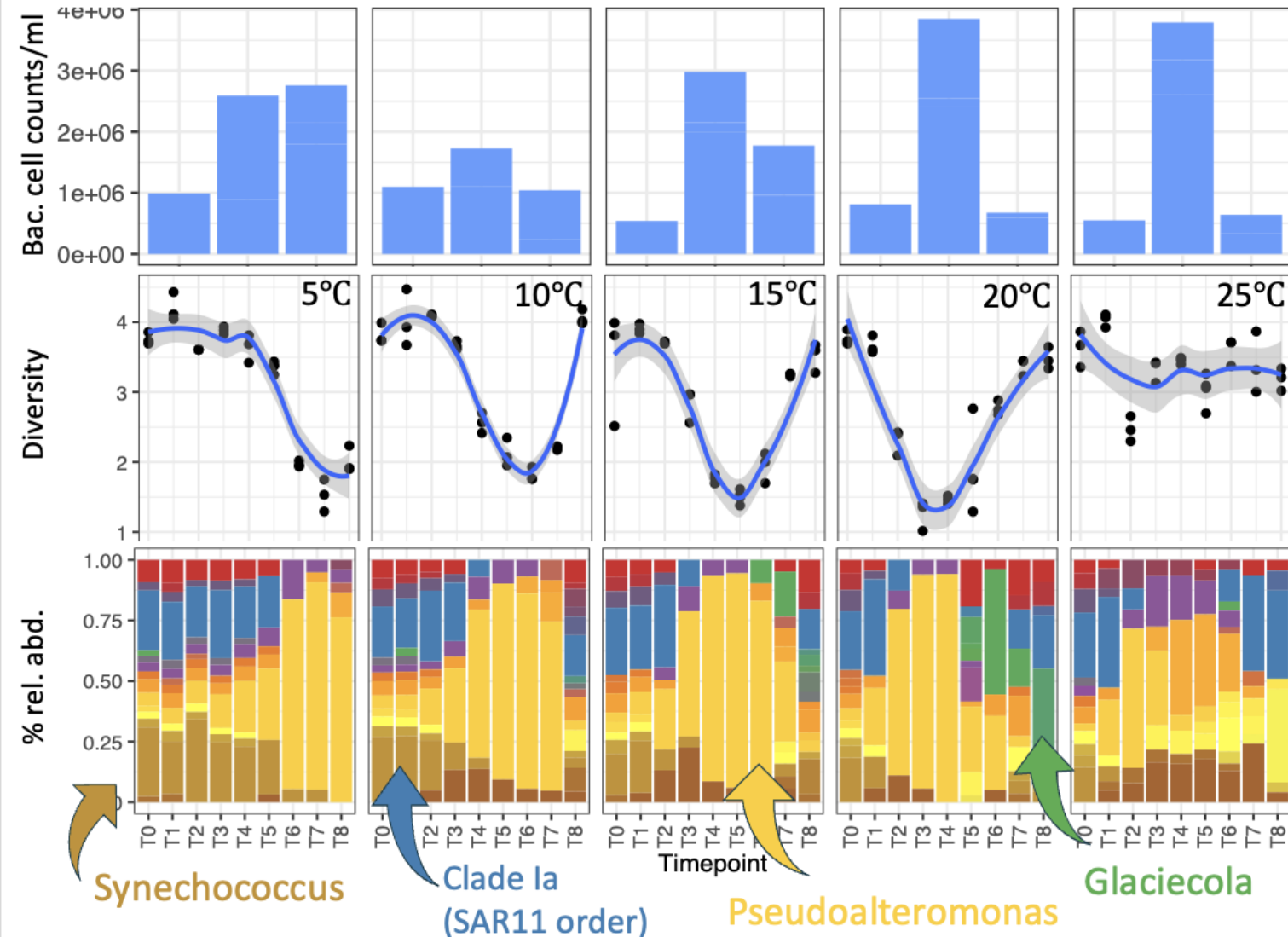


Microbial community changes over time - FISH



- Species richness drops when *Pseudoalteromonas* dominates the microbial community
- As temperature increases, the *Pseudoalteromonas* dominance occurs at **earlier timepoints**
- This **boom/bust cycle** was also apparent in bacterial cell counts

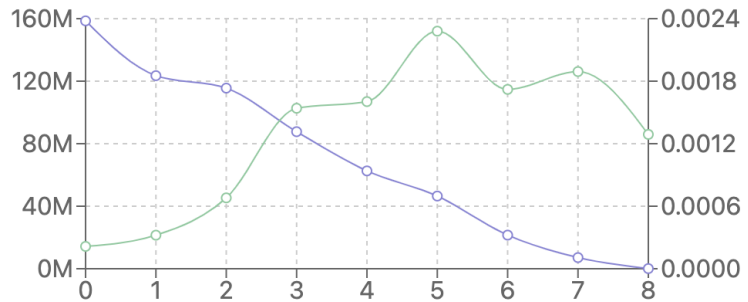
Microbial community changes over time - FISH



- Species richness drops when *Pseudoalteromonas* dominates the microbial community
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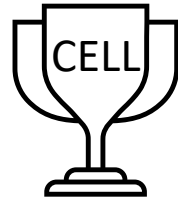
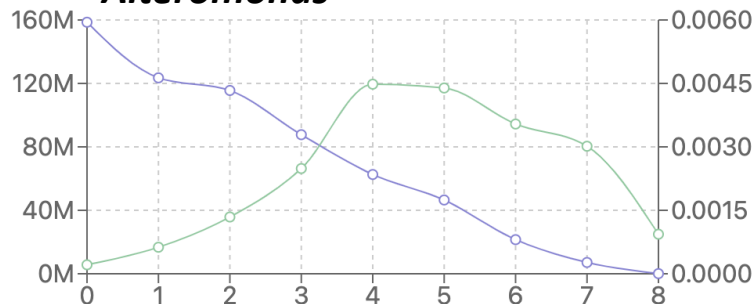
Putative marine eDNA degraders

Pseudoalteromonas



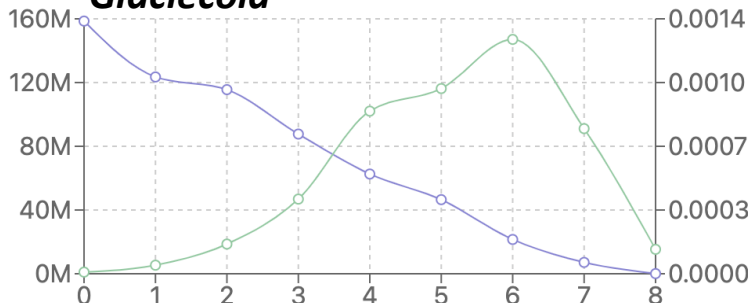
- Known for producing DNases that can break down extracellular DNA in marine environments
- **Often found in particle-attached communities**

Alteromonas



- Produces various hydrolytic enzymes and can nucleases for phosphorus acquisition
- Often found in **marine aggregates** where extracellular DNA is abundant

Glaciecola



- Adapted to utilizing various **dissolved organic matter** in marine environments
- Can produce extracellular enzymes for substrate degradation

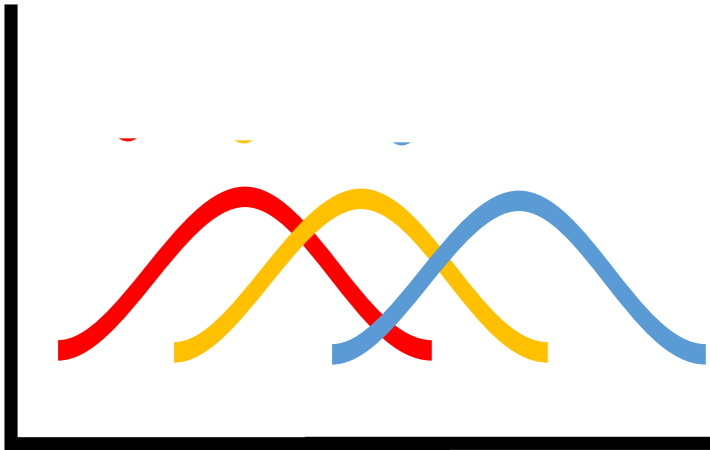
Fish gene copies (per 250 ml)

% relative abundance

Timepoint

Conclusions

Bacteria exhibited a boom-and-bust cycle of eDNA degradation



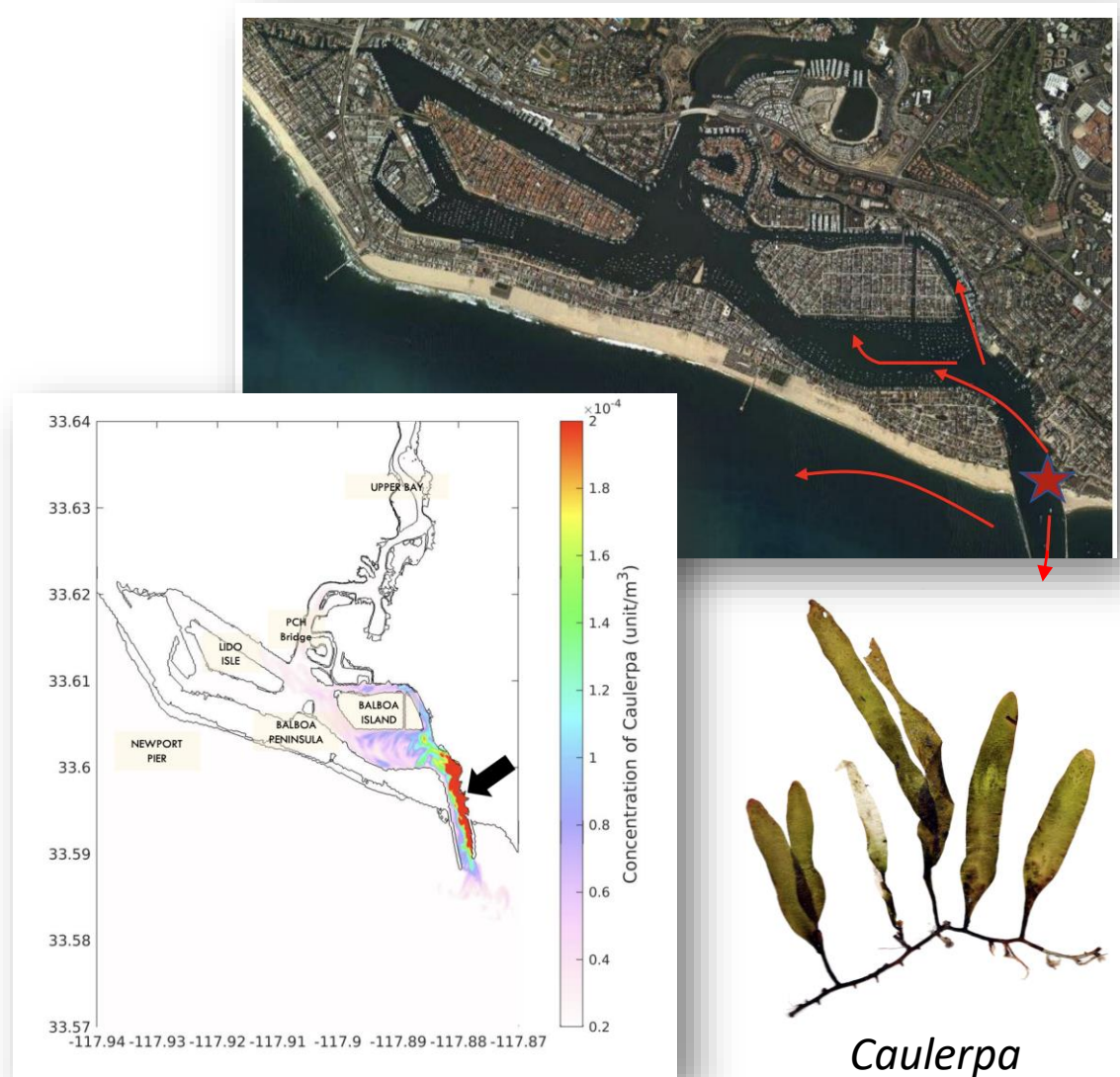
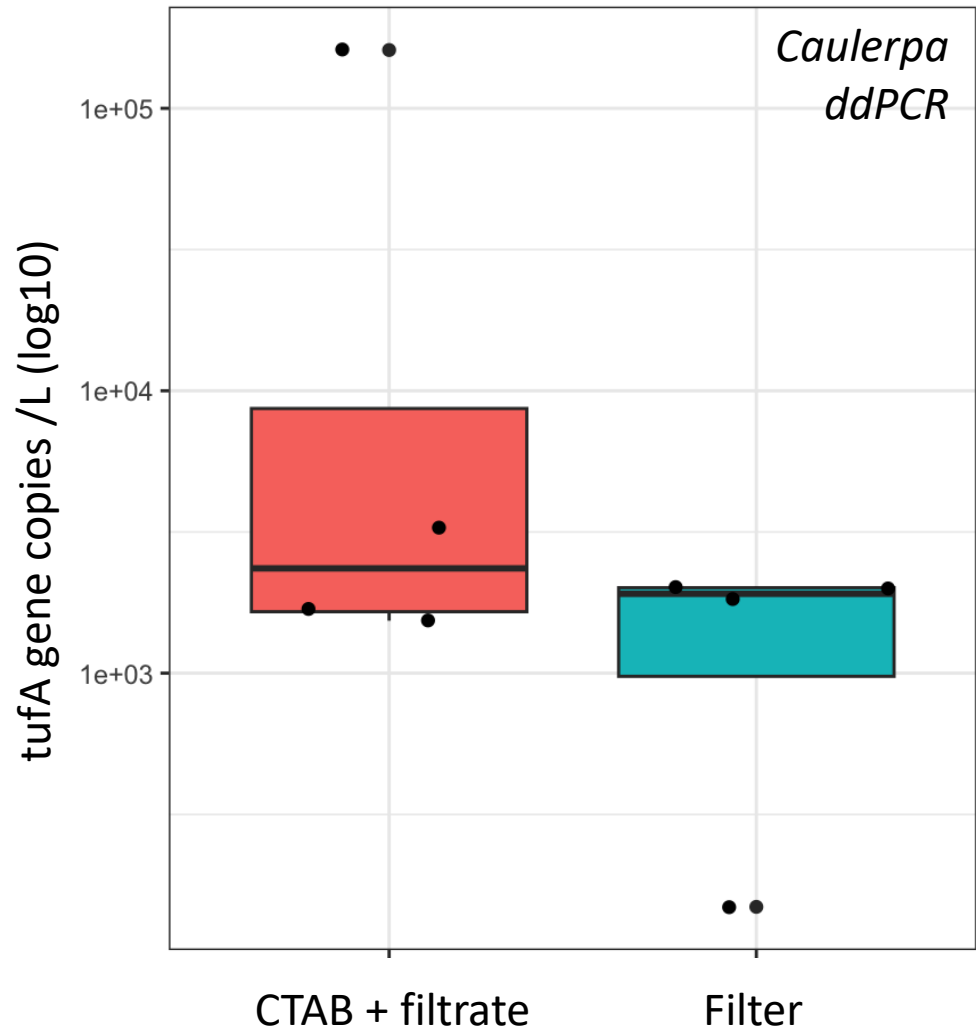
Bacterial “champion” degrader differed by eDNA state



Dissolved eDNA may persist longer than expected



Next steps: integrating decay rates into estuary models



Thank you! Contact: susannat@sccwrp.org



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Riley Darrough
USC



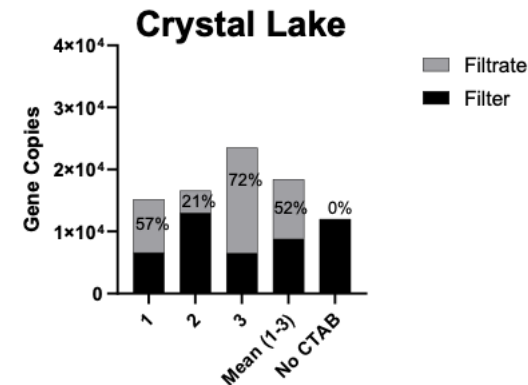
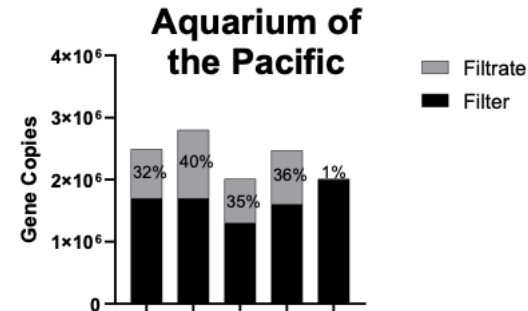
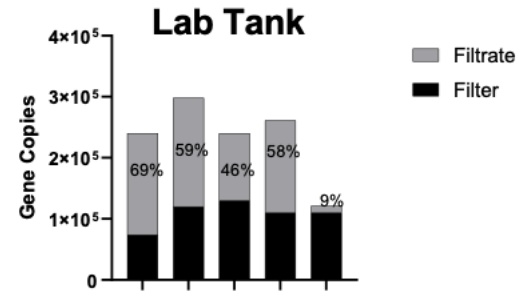
John Griffith,
SCCWRP

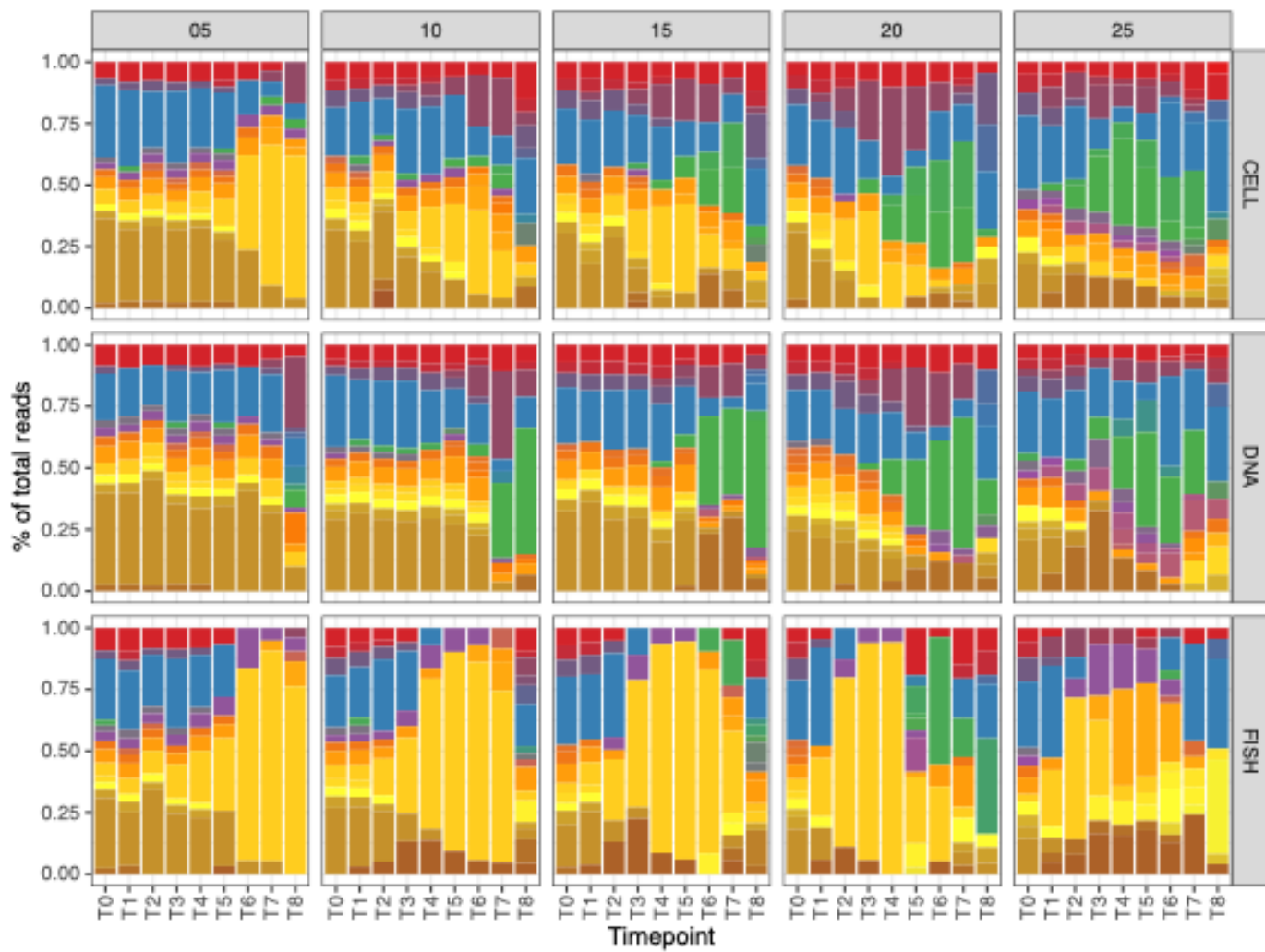


Next steps: what is in the dissolved DNA fraction?



~65% of Steelhead gene copies were in the dissolved fraction

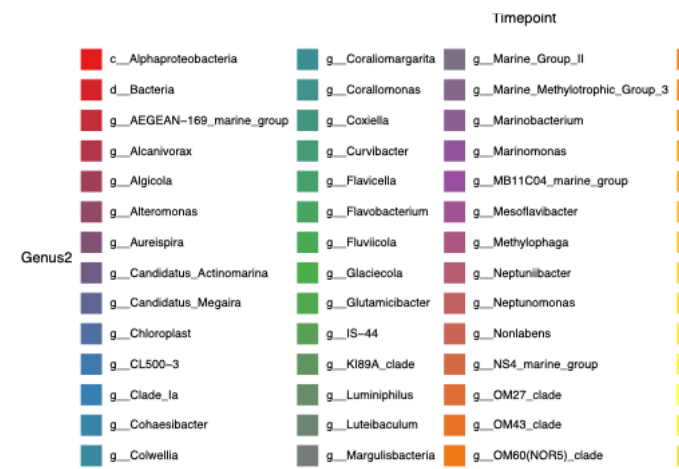




Alteromonas

Glaciecola

Pseudoalteromonas



Catalina mesocosm seawater

Time (UTC)	Temperature (°C)	Pressure (dbar)	Salinity (PSU)	DO (ml/l)	CHLA (µg/L)	Turbidity (NTU)	Volts	NO3 µM	NO2	PO4	SIL	NH4
4/28/24 20:27	16.2669	1.84	33.244	5.575	0.713	0.451	11.83	1.76	0.07	0.29	6.3	2.35
Sea-Bird Scientific Water Quality Monitor (WQM) sensor												
Wrigley dataset see here												

Sample ID	NO3 µmol/L	NO2 µmol/L	PO4 µmol/L	SIL µmol/L	NH4 µmol/L	Lab Temperature Celsius	Comment
Catalina SW	1.76	0.07	0.29	6.3	2.35	21.7	